

Quantification of ion binding using electrospray mass spectrometry†

Cite this: *Inorg. Chem. Front.*, 2014, **1**, 49

Received 10th September 2013,
Accepted 23rd October 2013

DOI: 10.1039/c3qi00037k

rsc.li/frontiers-inorganic

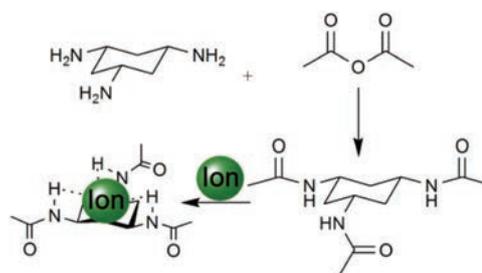
Jennifer S. Mathieson, Geoffrey J. T. Cooper, Mark D. Symes and Leroy Cronin*

Mass spectrometry has predominately been used as a qualitative tool in the analysis of complexes with quantitative studies being difficult to validate. Here, we present a method where electrospray mass spectrometry is used to give both a qualitative and quantitative measure of ion binding with cross validation by solution ^1H NMR, demonstrating it is possible to utilize gas-phase electrospray mass spectrometry continuous variation data to deduce binding stoichiometries in solutions where artifacts and nonspecific complexes are also observed.

The development of artificial synthetic receptors has been expanding greatly over the past decade^{1–4} as supramolecular principles are applied across an ever increasing range of molecules, materials, and systems.^{5–7} Over the past two decades analysis of such supramolecular complexes has become increasing straightforward with the utilization of soft ionization techniques, such as fast atom bombardment (FAB), matrix-assisted laser desorption (MALDI) and electrospray ionisation (ESI-MS)^{8–14} and studies using NMR methods such as the continuous method of variation (Job plot) and NMR titration experiments.^{15–20} In 2002, Miyake *et al.* used fast atom bombardment mass spectrometry (FAB-MS) to provide evidence of inclusion complex formation between a cyclophane and an aromatic guest by constructing a Job plot from the absolute intensity of the observed complexation.^{21,22} Also, Van Dorselaer in 1996 conducted dilution based ESI studies to provide insight into thermodynamic equilibrium in solution.²³ Surprisingly though, no method utilizing electrospray mass spectrometry (ESI-MS) has been developed to form Job plots. Given that ESI-MS is a soft ionisation technique, we hypothesized that it may be possible to probe the formation of certain coordination complexes with little perturbation. The nature of mass spectrometry is such that peaks from artifacts and nonspecific complexes may be observed along with the

thermodynamically preferred species and so a clear indication of the preferred stoichiometry of the complex is unattainable from peak observation alone. With this method the true stoichiometry can be found. To explore this, we set out to develop an experiment protocol that utilizes the method of continuous variation to evaluate the binding stoichiometries of three receptor–ion pairings and compare both NMR and ESI-MS methods to illustrate that ESI-MS can be used as a tool in the quantitative and qualitative analysis of certain labile coordination complexes. This method also allowed the study of the stoichiometry of paramagnetic complexes which, due to the broadening and overlapping of peaks, has proven difficult using conventional NMR techniques. It has also allowed low concentration studies removing some of the issues that could arise when carrying out quantitative studies at higher concentrations. The soft nature of the ionization may also allow the observation of weak interactions that may not be seen with more harsh conditions.

The first receptor that was examined was the receptor *cis*, *cis*-1,3,5-tris-acetyl-amino-cyclohexane **1** which was synthesised from a ligand that has been extensively studied, *cis*, *cis*-1,3,5-triaminocyclohexane (*cis*-tach),^{24–26} and acetic anhydride (see Scheme 1). As the aim of the research was to evaluate the use of ESI-MS as a quantitative analytical tool, the ligand was designed to be simple and also to be able to bind spherical anions such as chloride, bromide and iodide.



Scheme 1 A simple tripodal *cis*-tach anion receptor **1** synthesised from *cis*-tach and acetic anhydride refluxed with triethylamine in methanol for 24 hours.

University of Glasgow, School of Chemistry, Joseph Black Building, University Avenue, Glasgow, G12 8QQ, UK. E-mail: Lee.Cronin@glasgow.ac.uk

† Electronic supplementary information (ESI) available: Tables of ^1H NMR and mass spectral data. See DOI: 10.1039/c3qi00037k

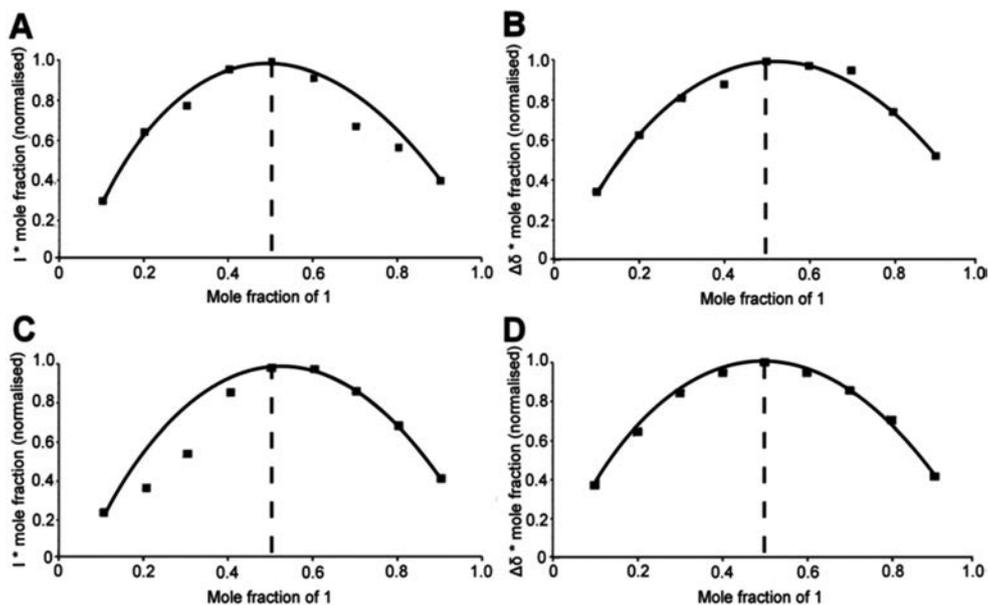


Fig. 1 (A) MS (CH_3OH) Job plot of **1** + Cl^- showing a 1 : 1 (1 : Cl^-) stoichiometry; (B) ^1H NMR (d_6 -DMSO) Job plot of **1** + Cl^- showing a 1 : 1 (1 : Cl^-) stoichiometry; (C) MS (CH_3OH) Job plot of **1** + Br^- showing a 1 : 1 (1 : Br^-) stoichiometry; (D) ^1H NMR (d_6 -DMSO) Job plot of **1** + Br^- showing a 1 : 1 (1 : Br^-) stoichiometry.

In order to obtain the stoichiometries of the **1**-anion complexes, an ESI-MS method was developed analogous to the method of continuous variation (or Job plots)²⁷ used with NMR where standard solutions (0.02 M) of **1** and the salt (KCl/KBr/KI) were made up in methanol and varying ratios of each solution were taken and mixed together (with the overall concentration remaining the same) *e.g.* 1 mL of **1** solution mixed with 9 mL of the salt solution, 2 mL of **1** solution mixed with 2 mL of the salt solution, 3 mL of **1** solution mixed with 7 mL of the salt solution up to 9 mL of **1** solution mixed with 1 mL of the salt solution. ESI-MS at 30 °C of the reaction mixtures was then carried out (where the acquisition parameters were kept the same for each experiment) and the intensity of the corresponding peak recorded. The recorded change in intensity multiplied by the mole fraction (where mole fraction is $[\text{Host}]/([\text{Host}] + [\text{Guest}])$) was then plotted against mole fraction to give a maximum value at 0.5 mole fraction and therefore a stoichiometry of 1 : 1 (1 : A) for each guest *e.g.* the Job plot of **1** with KCl in methanol was achieved by plotting the recorded change in intensity of the $[\text{C}_6\text{H}_{12}\text{N}_3(\text{C}_2\text{H}_3\text{O})_3\text{Cl}]^-$ (290.1 m/z) peak (see Fig. 2) multiplied by mole fraction ($[\text{Host}]/([\text{Host}] + [\text{Guest}])$)²⁷ against mole fraction to give a maximum at 0.5, indicative of a 1 : 1 stoichiometry (see Fig. 1, part A). It should be noted here that a small abundance of higher order stoichiometries were also present in the spectrum but applying the same method to these species did not give a continued variation and so we can postulate that this method will only work on the species with the correct thermodynamically preferred stoichiometry. This method was also applied to **1** in the presence of KBr (see Fig. 1 parts C and D) and KI to produce ESI-MS Job plots both with maxima at 0.5 (see ESI† for experimental details and MS parameters).

By using conventional ^1H NMR methods, the complexation of the receptor **1** to the three different anions Cl^- , Br^- and I^- was also studied and compared. The experimental set up was as before, with the mass spectrometry experiment where standard solutions (0.02 M) of **1** and the salt (KCl/KBr/KI) were made up in d_6 -DMSO. Varying ratios of each solution were then taken and mixed together (with the overall concentration remaining the same). ^1H NMR experiments were carried out for each of the mixtures and the receptor where a plot of $\Delta\delta$ (where $\Delta\delta$ = the shift of the receptor – (shift of the receptor + guest)) multiplied by the mole fraction was then plotted against mole fraction. It was observed that the NH chemical shifts of the receptor shifted upfield in the presence of the anionic guest, indicating that complexation had occurred (see Fig. 3). A Job plot was then produced for each anion with the maximum at 0.5 for each, corresponding to a stoichiometry of 1 : 1 (see Fig. 1 parts B and D, Fig. 3 and the ESI†).

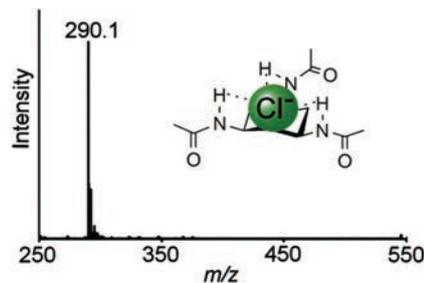


Fig. 2 Mass spectrum at 30 °C of the receptor **1** in the presence of KCl giving the species $[\text{C}_6\text{H}_{12}\text{N}_3(\text{C}_2\text{H}_3\text{O})_3\text{Cl}]^-$ (290.1 m/z).

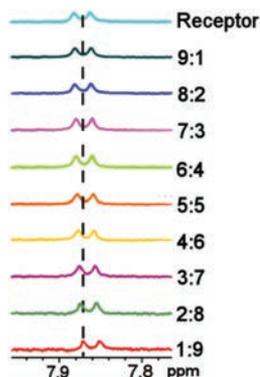


Fig. 3 ^1H NMR (d_6 -DMSO) showing the shift of the amide NH when hydrogen bonding to chloride occurs; ratio is receptor to anion.

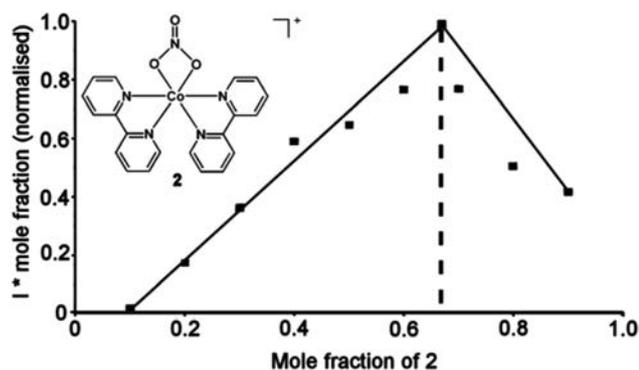


Fig. 4 Reaction scheme and Job plot of $[\text{Co}(\text{C}_{10}\text{H}_8\text{N}_2)_2(\text{NO}_3)]^- 2$ in the presence of $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ in methanol at 30°C showing a 2:1 receptor : metal stoichiometry.

Job plot studies of 12-crown-4 with Na^+ and Li^+ were then carried out using ESI-MS and ^1H NMR to demonstrate the observation of more complex stoichiometries. Both ESI-MS and ^1H NMR job plots showed a 1:1 (12-crown-4 : cation) stoichiometry when Li^+ was coordinated to the crown ether and a stoichiometry of 2:1 (12-crown-4 : Na) was found with both methods when Na^+ was coordinated (see ESI† for the Job plots of 12-crown-4 with Na^+ and Li^+).

The method developed for the mass spectrometry experiments was then applied to a complex whose stoichiometry could not be obtained using ^1H NMR methods due to its paramagnetic behavior. A Job plot of the complex $[\text{Co}(\text{C}_{10}\text{H}_8\text{N}_2)_2(\text{NO}_3)]^- 2$ (433.1 m/z) (see Fig. 4), which was formed when 2,2'-bipyridine was reacted in the presence of $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ in methanol, was produced where the maximum of the normalized intensity was at 0.67 mole fraction therefore showing a 2:1 receptor to metal stoichiometry (see Fig. 4).

Conclusions

In conclusion, this research has shown that mass spectrometry, in the case of the above coordination complexes, can be used as both a qualitative and quantitative analytical tool.

It can be used to demonstrate binding stoichiometries not only through observation of a species, but also through a continuous variation method. Often, with the conventional ^1H NMR method, confirmation of complex formation can be problematic with broadening and overlapping of peaks, and so the use of ESI-MS to analyse the data removes some of the ambiguity that comes with using NMR methods.

Notes and references

- N. Busschaert, I. L. Kirby, S. Young, S. J. Coles, P. N. Horton, M. E. Light and P. A. Gale, *Angew. Chem., Int. Ed.*, 2012, **51**, 4426.
- (a) C. Caltagirone, J. R. Hiscock, M. B. Hursthouse, M. E. Light and P. A. Gale, *Chem.-Eur. J.*, 2008, **14**, 10236; (b) C. Lincheneau, R. M. Duke and T. Gunnlaugsson, *Org. Biomol. Chem.*, 2012, **10**, 6069.
- S. Comby, S. A. Tuck, L. K. Truman, O. Kotova and T. Gunnlaugsson, *Inorg. Chem.*, 2012, **51**, 10158.
- (a) P. V. Mason, N. R. Champness, S. R. Collinson, M. G. Fisher and G. Goretzki, *Eur. J. Org. Chem.*, 2006, 1444; (b) S. K. Kim, D. E. Gross, D.-G. Cho, V. M. Lynch and J. L. Sessler, *J. Org. Chem.*, 2011, **76**, 1005; (c) S. K. Kim, B. P. Hay, J. S. Kim, B. A. Moyer and J. L. Sessler, *Chem. Commun.*, 2013, **49**, 2122; (d) S. O. Kang, R. A. Begum and K. Bowman-James, *Angew. Chem., Int. Ed.*, 2006, **45**, 7882.
- (a) A. C. Sather, O. B. Berryman and J. Rebek Jr., *J. Am. Chem. Soc.*, 2010, **132**, 13572; (b) P. M. P. Anderson, R. J. Gregory, S. Thompson, D. W. Souza, S. Paul, R. C. Mulligan, A. E. Smith and M. J. Welsh, *Science*, 1991, **253**, 202.
- (a) Y. Yang, B. Szamosfalvi, J. Yee, S. Frinak and E. V. Anslyn, *Analyst*, 2011, **136**, 317; (b) P. A. Gale, *Coord. Chem. Rev.*, 2003, **240**, 191; (c) P. D. Beer and P. A. Gale, *Angew. Chem., Int. Ed.*, 2001, **40**, 486.
- (a) S. K. Kim, G. I. Vargas-Zúñiga, B. P. Hay, N. J. Young, L. H. Delmau, C. Masselin, C.-H. Lee, J. S. Kim, V. M. Lynch, B. A. Moyer and J. L. Sessler, *J. Am. Chem. Soc.*, 2012, **134**, 1782; (b) J. J. Gassensmith, H. Furukawa, R. A. Smaldone, R. S. Forgan, Y. Y. Botros, O. M. Yaghi and J. F. Stoddart, *J. Am. Chem. Soc.*, 2011, **133**, 15312; (c) E. Busseron and J. Rebek Jr., *Org. Lett.*, 2010, **12**, 4828.
- C. A. Schalley, *Int. J. Mass Spectrom.*, 2000, **194**, 11.
- (a) H. Zhang, M. Grabenauer, M. T. Bowers and D. V. Dearden, *J. Phys. Chem. A*, 2009, **113**, 1508; (b) D. V. Dearden, T. A. Ferrell, M. C. Asplund, L. W. Zilch, R. R. Julian and M. F. Jarrold, *J. Phys. Chem. A*, 2009, **113**, 989; (c) C. A. Schalley, J. Hoernschemeyer, X. Li, G. Silva and P. Weis, *Int. J. Mass Spectrom.*, 2003, **228**, 373; (d) C. A. Schalley, T. Martín, U. Obst and J. Rebek Jr., *J. Am. Chem. Soc.*, 1999, **121**, 2133; (e) T. Becherer, D. Meshcheryakov, A. Springer, V. Böhmer and C. A. Schalley, *J. Mass Spectrom.*, 2009, **44**, 1338; (f) T. Shiga, G. N. Newton, J. S. Mathieson, T. Tetsuka, M. Nihei, L. Cronin and H. Oshio, *Dalton Trans.*, 2010, **39**, 4730.

- 10 (a) S. E. Pierce, R. Kiełtyka, H. F. Sleiman and J. S. Brodbelt, *Biopolymers*, 2008, **91**, 233; (b) J. B. Shaw, J. A. Madsen, H. Xu and J. S. Brodbelt, *J. Am. Soc. Mass Spectrom.*, 2012, **23**, 1707; (c) S. I. Smith, F. S. Guziec Jr., L. Guziec and J. S. Brodbelt, *Analyst*, 2009, **134**, 2058; (d) J. S. Brodbelt, *Annu. Rev. Anal. Chem.*, 2010, **3**, 67; (e) C. L. Mazzitelli, Y. Chu, J. J. Reczek, B. L. Iverson and J. S. Brodbelt, *J. Am. Soc. Mass Spectrom.*, 2007, **18**, 311.
- 11 D.-L. Long, C. Streb, Y.-F. Song, S. Mitchell and L. Cronin, *J. Am. Chem. Soc.*, 2008, **130**, 1830.
- 12 (a) S. Fukuzumi, K. Ohkubo, D. S. Kim, J. S. Park, A. Jana, V. M. Lynch, D. Kim and J. L. Sessler, *J. Am. Chem. Soc.*, 2011, **133**, 15938; (b) J. S. Park, E. Karnas, K. Ohkubo, P. Chen, K. M. Kadish, S. Fukuzumi, C. W. Bielawski, T. W. Hudnall, V. M. Lynch and J. L. Sessler, *Science*, 2010, **329**, 1324; (c) K. Tiefenbacher and J. Rebek Jr., *J. Am. Chem. Soc.*, 2012, **134**, 2914; (d) F. Cui, S. Li, C. Jia, J. S. Mathieson, L. Cronin, X.-J. Yang and B. Wu, *Inorg. Chem.*, 2012, **51**, 179; (e) A. Coskun, M. Hmadeh, G. Barin, F. Gándara, E. Choi, N. L. Strutt, D. B. Cordes, A. M. Z. Slawin, J. F. Stoddart, P. Sauvage and O. M. Yaghi, *Angew. Chem., Int. Ed.*, 2012, **51**, 2160; (f) J.-F. Ayme, J. E. Beves, D. A. Leigh, R. T. McBurney, K. Rissanen and D. Schultz, *Nat. Chem.*, 2012, **4**, 15; (g) S. K. Kim, J. L. Sessler, D. E. Gross, C.-H. Lee, J. S. Kim, V. M. Lynch, L. H. Delmau and B. P. Hay, *J. Am. Chem. Soc.*, 2010, **132**, 5827.
- 13 L. Vilà-Nadal, A. Rodríguez-Forteza, L.-K. Yan, E. F. Wilson, L. Cronin and J. M. Poblet, *Angew. Chem., Int. Ed.*, 2009, **48**, 144.
- 14 A. V. Simionato, M. D. Cantú and E. Carrilho, *Microchem. J.*, 2006, **82**, 214.
- 15 D. K. Smith, *Org. Biomol. Chem.*, 2003, **1**, 3874.
- 16 P. D. Beer, C. Hazelwood, D. Heseck, J. Hodacova and S. E. Stokes, *Dalton Trans.*, 1993, 1327.
- 17 P. A. Gale and T. Gunnlaugsson, *Chem. Soc. Rev.*, 2010, **39**, 3595.
- 18 C. Markert and A. Pfaltz, *Angew. Chem., Int. Ed.*, 2004, **43**, 2498.
- 19 D. Feichtinger and D. A. Plattner, *Angew. Chem., Int. Ed.*, 1997, **36**, 1718.
- 20 M. T. Blanda, J. H. Horner and M. Newcom, *J. Org. Chem.*, 1989, **54**, 4626.
- 21 M. Koichi, Y. Kimura and M. Miyake, *J. Mass Spectrom. Soc. Jpn.*, 2002, **50**, 301.
- 22 M. Koichi, Y. Kimura and M. Miyake, *J. Mass Spectrom. Soc. Jpn.*, 2003, **51**, 566.
- 23 E. Leize, A. Jaffrezic and A. Van Dorsselaer, *J. Mass Spectrom.*, 1996, **31**, 537.
- 24 A. L. Pickering, G. Seeber, D.-L. Long and L. Cronin, *Chem. Commun.*, 2004, 136.
- 25 G. N. Newton, G. J. T. Cooper, D. Schuch, T. Shiga, S. Khanra, D.-L. Long, H. Oshio and L. Cronin, *Dalton Trans.*, 2009, 1549.
- 26 A. L. Pickering, G. J. T. Cooper, D.-L. Long and L. Cronin, *Polyhedron*, 2004, **23**, 2075.
- 27 P. Job, *Ann. Chim. Appl.*, 1928, 113.
- 28 R. Ivaníková, I. Svoboda, A. Mašlejová, B. Papánková and H. Fuess, *Acta Crystallogr., Sect. E: Struct. Rep. Online*, 2006, **62**, 3024.